Storage stability of bevacizumab in polycarbonate and polypropylene syringes Hanieh Khalili^{1,2}*, Garima Sharma^{1,2}°, Andrew Froome³, Peng Tee Khaw¹, Steve Brocchini^{1,2} ¹NIHR Biomedical Research Centre, Moorfields Eye Hospital and UCL Institute of Ophthalmology, London, EC1V 9EL, UK ²UCL School of Pharmacy, University College London, 29-39 Brunswick Square, London WC1N 1AX, UK ³Moorfields Pharmaceuticals (Moorfields Eye Hospital), 34 Nile Street, London, N1 7TP, UK *Corresponding Author: <u>Hanieh.khalili@ucl.ac.uk</u> [°] Equal contribution for first authorship

Abstract

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37 Purpose To compare and examine the storage stability of compounded bevacizumab in polycarbonate (PC) and polypropylene (PP) syringes over a 6-month period. PC 38 39 syringes have been used in a recent clinical study and bevacizumab stability has not 40 been reported for this type of syringe. 41 Methods Repackaged bevacizumab was obtained from Moorfields Pharmaceuticals in 42 polycarbonate (PC) and polypropylene (PP) syringes. Bevacizumab from the stored 43 syringes was analysed at monthly time points for a 6-month period and compared 44 with bevacizumab from a freshly opened vial at each time point. SDS-PAGE 45 electrophoresis and size-exclusion chromatography (SEC) was used to observe 46 aggregation and degradation. Dynamic light scattering (DLS) provided information 47 about the hydrodynamic size and particle size distribution of bevacizumab in solution. 48 VEGF binding and the active concentration of bevacizumab was determined by 49 surface plasmon resonance (SPR) using Biacore. 50 Results SDS-PAGE and SEC analysis did not show any changes in the presence of 51 higher molecular species (HMWS) or degradation products in PC and PP syringes 52 from T0 to T6 compared to bevacizumab sampled from a freshly opened vial. The 53 hydrodynamic diameter of bevacizumab in the PC syringe after six months of storage 54 was not significantly different to bevacizumab taken from a freshly opened vial. 55 Using SPR, the VEGF binding activity of bevacizumab in the PC syringe was 56 comparable with bevacizumab taken from a freshly opened vial. 57 Conclusion No significant difference over a 6-month period was observed in the 58 quality of bevacizumab repackaged into prefilled PC polycarbonate and PP 59 polypropylene syringes when compared to bevacizumab that is supplied from the vial. 60 61 Keywords; bevacizumab, compounded bevacizumab, storage stability 62 63 64

Introduction

Two recent multi-center randomised controlled clinical trials compared the use of ranibizumab (Lucentis, Genentech) and bevacizumab (Avastin, Genentech) to treat wet age-related macular degeneration (AMD).¹⁻⁴ These trials (IVAN and CATT) found there is no difference in visual acuity outcome during one and two year treatment periods respectively.^{2, 3} Both ranibizumab and bevacizumab were developed to bind to vascular endothelial growth factor (VEGF) as a means to inhibit blood vessel growth.⁵ Ranibizumab is a humanised antibody fragment (Fab) that is licensed for intravitreal injection to treat AMD and other retinal conditions.⁶⁻¹⁰

Bevacizumab is a humanised monoclonal full-length antibody that is licensed for administeration by intravenous infusion to treat cancer (metastatic colorectal, NSCLC, renal cell cancer, glioblastoma).^{11, 12} It is not licensed for intravitreal injection to treat retinal diseases. Bevacizumab is normally provided as a solution in a glass vial containing 400 mg of the antibody at a concentration of 25 mg/mL. For ocular use, bevacizumab is often transferred under aseptic conditions into ready-to-use 1.0 ml syringes for intravitreal injection by compounding pharmacies for local distribution. A shelf life of up to 3 months^{13, 14} is often specified. To avoid the risks and costs of compounding there have been reports of 'multiple use' from a vial of bevacizumab to treat patients consecutively. However there is the risk of infection if the vial is punctured multiple times and an increased incidence of endophthalmitis has been reported.¹⁵

The National Institute for Clinical—Health and Care Excellence (NICE) considers the compounding of bevacizumab into syringes followed by storage prior to ophthalmic use to be unlicensed, rather than off-license use of bevacizumab. ¹⁶ In spite of head-to-head trials indicating that ranibizumab and bevacizumab are clinically statistically equivalent, some safety results from the CATT study indicate there may be a greater burden of side effects for bevacizumab compared to ranibizumab.

The cost of <u>compounded</u> bevacizumab per intravitreal dose is approximately 5-9% of the cost of a dose of ranibizumab.¹⁷ Moderate to severe disabilities in our ageing population, of which diminishing visual function is one, are projected to increase by 32-54% in the UK by 2022.¹⁷ Ranibizumab and bevacizumab are used for other major ophthalmic diseases affecting older patients including diabetic retinopathy and retinal vein occlusion, while AMD is the main cause of blindness for

these older patients.¹⁸ Unfortunately costs have generally become a constraining factor for the use of expensive medicines in many parts of the world. It is not unreasonable to expect that intravitreal use of bevacizumab will continue in many parts of the world, especially in resource limited regions and especially for older patients whose overall health and social care costs are already high and are expected to increase.^{17, 18}

The reported incidence of IOP spike^{19, 20} or endophthalmitis that may be associated with bevacizumab injections²¹⁻²⁴ is thought to be related to the presence of particulates or protein aggregates.^{20, 25} The presence of silicon oil contamination and the type of syringe used for repackaged bevacizumab has also been reported to be associated with an increase in protein aggregates or particulate count.¹⁴ As with any protein therapeutic antibodies, exposure to light, storage temperature, product handling, and syringe components can cause protein misfolding, denaturation and aggregation. These changes in protein structure can decrease protein activity and may result in immunological responses.²⁶

Ranibizumab has recently become available in ready-to-use glass syringes^{27, 28} but the cost of this medicine has yet to drop. Unfortunately the compounding and subsequent storage of bevacizumab in plastic syringes have not been approved by regulatory agencies. One important factor to consider is the different types of syringe that are used for bevacizumab. Reports have been published about bevacizumab being compounded into polypropylene (PP) syringes^{13, 14, 29, 30} and the effects of storage conditions^{20, 31} on the stability and efficacy of bevacizumab. Polycarbonate (PC) syringes have also been used, and were used in the IVAN trial.^{20, 32} There does not however appear to be a report about the stability of bevacizumab when repackaged in polycarbonate syringes (Table 1).

124 TABLE 1

In this study, we examined the stability of compounded bevacizumab in both polycarbonate (PC) and polypropylene (PP) syringes. The PC syringe had a luer-lock to secure the needle. The more commonly used PP syringe had a slip-lock to hold the needle. The bevacizumab filled syringes were then stored at 5 ± 3 °C. The stored bevacizumab filled syringes were evaluated monthly over a 6-month period by SDS-PAGE gel electrophoresis, size-exclusion chromatography (SEC), dynamic light scattering (DLS), and surface plasmon resonance (SPR).

Materials and Methods

134 Materials

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135 Bevacizumab (Avastin, Genentech, 400 mg) solution from a vial (16 mL) was aseptically fractionated into 1.0 mL sterile syringes at Moorfields Pharmaceuticals a 136 137 day before starting the first time point. The volume of the bevacizumab solution 138 transferred to each syringe was 0.13 mL. Two different syringes (as shown in table 2) 139 were evaluated, polycarbonate (PPPC) with a luer-lock barrel and (ii) polypropylene 140 (PP) with a slip-lock barrel (Table 2). A fresh vial of bevacizumab was used for the 141 control data obtained at each time point. The filled syringes and vials were stored at 142 5° C $\pm 3^{\circ}$ C and the temperature was monitored and recorded at regular intervals at 143 Moorfields Pharmaceuticals. At each sampling time point, syringes were shipped to 144 the UCL School of Pharmacy where stability studies were conducted within 24 hour 145 of receipt.

TABLE 2

Methods

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- 148 Study design
- 149 The bevacizumab solution in PC and PP syringes was evaluated for its
- physicochemical stability over a 6-month period at monthly time points and compared
- with bevacizumab solution obtained from freshly opened vials. Only one vial was
- used per time point and data generated during 8 hour period on the day that vial was
- open. The time points are designated as T0 (first time point after fractionation
- procedure), T3 and T6 representing three and six-month storage period.
- 155 Gel-electrophoresis SDS-PAGE
- Novex bis-tris 4-12% precast gels (Invitrogen, UK) were used for PAGE analysis.
- Solutions were first prepared by taking the bevacizumab solution from a syringe (0.05)
- mL) and the same volume of bevacizumab solution from the vial (0.05 mL) and
- adding each to Phosphate Buffered Saline (PBS), pH 7.2 to make up the volume to
- 1.0 mL and giving a final concentration of 1.25 mg/mL. PBS was prepared with
- tablets purchased from Oxoid, UK containing 0.16 M NaCl, 0.003 M KCl, 0.008 M
- Na₂HPO₄ and 0.001 M KH₂PO₄. Samples were then loaded (0.01 mL) onto a gel after
- mixing with SDS sample buffer (×4). Gels were then stained with Instant blue
- 164 (Expedeon Ltd, UK) staining to visualize the protein lane.

- 165 Size-exclusion chromatography
- 166 For SEC analysis the bevacizumab solution from a freshly open vial and the stored
- syringes was diluted with PBS (1.25 mg/mL, 1.0 mL) and transferred to sample vials
- in an autosampler which then loaded 950 µL of each sample onto a SEC column,
- 169 (HiLoad 16/600 Superdex 200 prep grade column, GE Healthcare Life sciences, UK)
- for separation. SECs were conducted in triplicate for each time point for both syringe
- and vial samples using a system comprised of a UV detector (Jasco UV-1570, at 280
- nm) and HPLC pump (Jasco PU-980 Intelligent). Azur software was used to process
- 173 chromatographic data.
- 174 Dynamic light scattering
- 175 Malvern Zetasizer Nano-ZS, UK with 633 nm laser source was used for measuring
- 176 hydrodynamic diameter of bevacizumab. Contaminating particles such as dust in a
- solution can be detected in DLS and cause interference. Hence, bevacizumab solution
- 178 from vial and syringe was diluted with 0.22 µm filtered PBS to make 1.25 mg/mL
- solution. These were then subjected to measurement by DLS in 1.0 mL disposable
- 180 polystyrene cuvettes. Nano-ZS analysis software was used to analyze the
- measurements. Each measurement was an average of 25 runs of 10 seconds each,
- carried out in duplicate. DLS analysis was performed at time points (T0, T3 and T6)
- for the bevacizumab solution obtained from a freshly opened vial and from the
- syringe at each time point. Samples from 6 different PC syringes were evaluated at
- each time point and three samples were made from the vial.
- 186 Active protein concentration using Biacore
- Human recombinant VEGF₁₆₅ (38 kDa MW, purchased from Sigma Aldrich) was
- immobilised on a CM5 (534 RU). The high immobilisation level was selected for
- concentration assays. The immobilisation was performed using standard carbodiimide
- mediated coupling (NHS/EDC, 50/50) and ethanolamine (pH 8.5). Samples were
- prepared in HBS-EP running buffer (10 mM HEPES, pH 7.4, 150 mm NaCl, 3.0 mM
- 192 EDTA, 0.005% surfactant P20). All binding and concentration measurements were
- 193 conducted at 25°C at a flow rate of 10 µL/min. Chip regeneration was accomplished
- by exposure to 10.0 mM glycine–HCl (pH 1.5) for 1200 sec. Double-referencing was
- performed to account for bulk effects caused by changes in the buffer composition or
- nonspecific binding. Data were evaluated with BIAevaluation software (version 2.1)
- 197 in Biacore X-100.

198 Statistical Analysis 199 Data was analysed for statistical significance using Student's t-test and p value of 200 <0.05 was considered statistically different. Data is presented as mean \pm S.D. for at 201 least triplicate observations. 202 **Results** 203 *Gel-electrophoresis (SDS-PAGE)* 204 SDS-PAGE analysis was conducted by diluting bevacizumab solution from a syringe 205 (PC and PP, 0.05 mL) and the same volume of bevacizumab solution from the vial 206 (0.05 mL) with PBS buffer (0.95 mL, pH 7.4). Samples (0.01 mL, 1.25 mg/mL) were 207 analysed by SDS-PAGE (Figure 1A). Three individual samples each from the syringe 208 and the vial were evaluated. The band at 150 kDa in Figure 1A is the monomer of 209 bevacizumab. The gels were heavily and equally loaded in an effort to observe any 210 changes in the presence of higher molecular species (HMWS) of bevacizumab or 211 degradation products. No change in SDS-PAGE from T0 to T6 was observed for any 212 of the samples. 213 FIGURE 1A 214 *Size-exclusion chromatography* 215 Size-exclusion chromatography (SEC) was used in an effort to observe if there was 216 any aggregation of bevacizumab. Six replicates were obtained for samples stored in 217 syringes at each time point. Three replicates were obtained at each time point for the 218 samples obtained directly from freshly opened vials. A representative chromatogram 219 (Figure 1B) shows the HMWS of bevacizumab at a retention time of 59 min and 220 monomer at 72 min for bevacizumab stored in the PC syringe for 6 months. Figure 1C 221 is the area under the curve (AUC) for the HMWS of bevacizumab at different time points (T0, T3, T6) for PP and PC syringes as compared to the vial. There appeared to 222 223 be no significant change in the AUC of this HMWS over the 6-month period for the 224 PP and PC syringe stored samples as compared to the vial. 225 FIGURE 1B and C

226 Dynamic light scattering

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On examination with SDS PAGE and SEC, there was no difference in the physical stability of bevacizumab stored in either polycarbonate (PP) or polypropylene (PC) syringes. The result obtained for physical stability of bevacizumab fractionated in

polypropylene syringes was found to be in excellent agreement with a previously published extensive report.³⁰ Hence, we decided to focus on the polycarbonate syringes for further analysis as these were used for IVAN study and reports were submitted to MHRA without published public records.³²

The hydrodynamic diameter of bevacizumab stored in PC syringes was found to be 11.19 with PdI of 0.02 at 25°C (Figure 1D). There was no significant difference in the size distribution of bevacizumab stored in the syringe after six months of storage compared to bevacizumab taken fresh from the vial (Figure 1E).

FIGURE 1D and E

Surface plasmon resonance (SPR)

Binding of bevacizumab was evaluated by SPR (Biacore) to calculate the active concentration of bevacizumab in the syringe compared with the vial. If the binding of bevacizumab to VEGF decreases due to storage in a syringe, then differences in the active concentration of bevacizumab from a freshly opened vial should be apparent when evaluated by SPR.³³ The binding of bevacizumab fractionated into PC syringe was studied during 6-month storage period and compared to bevacizumab in freshly opened vial.

A CM5 chip was functionalized with VEGF (534 RU) to conduct the binding assay. The binding study was then performed on bevacizumab that was aliquoted from a freshly opened vial and from the syringe at each time point. Figure 2A shows a representative bar chart of the binding for bevacizumab in two concentrations (1.25 and 0.625 mg/mL) from a PC syringe after 6 months storage and from a freshly opened vial. There did not appear to be any difference in the binding response of bevacizumab from the syringe at the different time points compared to the bevacizumab from a freshly opened vial (Figure 2A). For reference, a superposition is shown in Figure 2B of the sensograms for the bevacizumab from both the vial and the syringe at the 6 month time point.

FIGURE 2A and B

To calculate the active concentration of bevacizumab from the syringe, a calibration standard curve was generated with bevacizumab (2.0-0.25 mg/mL, 4 dilutions) obtained from a freshly opened vial. Bevacizumab (1.25 mg/mL) from the PC syringe was prepared from 0.05 mL protein solution into 1.0 mL buffer and was passed over the CM5 chip to interact with immobilised VEGF. In parallel, 0.05 mL of

bevacizumab from the vial (1.25 mg/mL) was added to 1.0 mL of buffer and evaluated. The calibration responses were then used to calculate the active concentration of bevacizumab in the syringe and vial (Figure 2C). The amount of bevacizumab in the syringe did not change significantly compared to that observed for the vial and no difference was observed at T6 compared with T0.

268 FIGURE 2C

Discussion

There have been previous studies to investigate the physical stability of repackaged bevacizumab in polypropylene syringes.^{13, 14, 30} The results from our study suggest that there is no significant difference in the physical stability of bevacizumab when repackaged in polycarbonate or polypropylene syringes when compared to bevacizumab that had been stored in a glass vial. The study was performed at time points over a six-month period using different techniques.

Bevacizumab is a 150 kDa protein which is displayed as a distinct band (Figure 1A, Lanes 1-9) by SDS PAGE. There is a faint band seen at about the 260 kDa protein standard band. This may indicate the presence of aggregates of the antibody, which is also consistent with what was observed by SEC³⁰ (Figure 1B, peak at 59 min) with the presence of higher molecular weight species. To investigate this further, fractions were collected from 58-59 min and analysed with SDS-PAGE (Figure 3) using silver-stain as a detection method. Silver stain is more sensitive than colloidal blue staining and can detect protein in the range of 5-30 ng. ³⁴ Fractions were also collected at the main peak (71-72 min; Figure 3, Lanes 5-6). The higher molecular weight band was not observed at the peak of 71-72 min (Figure 3, Lanes 5-6) suggesting that this species was not in equilibrium with bevacizumab. The fractions obtained from the peak at 58-60 minutes appear to be a heterogeneous population with bevacizumab HMWS. This higher molecular weight band in SDS-PAGE has been reported previously¹³ for bevacizumab in vial and syringe. However, it is important to note that there is lack of significant difference in the level of HMWS between vial and PC syringe at different time points between T0 and T6.

FIGURE 3

There was no significant difference in the hydrodynamic radius of bevacizumab from the vial and the PC syringe over a six-month period when measured by DLS. A similar result was reported by Paul et al³⁰ for storage stability of

bevacizumab fractionated in a PP syringe for a period of three months. Paul et al³⁰ also reported that the high molecular weight species (HMWS) present in the bevacizumab solution in the PP syringe was approximately 360 nm when the DLS measurement was made at 25°C. However, the hydrodynamic size of the bevacizumab sample stored at ambient temperature overnight was 100.5 nm (PDI = 0.46) suggesting that the storage temperature has an impact on the bevacizumab stability profile.

SPR was used to evaluate bevacizumab binding to VEGF and no change in binding was observed during the 6-month storage period for the bevacizumab stored in the PC syringes compared to bevacizumab from the vial. SPR is a non-labeling technique that allows measurement of protein-protein interactions such as antibodyantigen interactions. One of the interacting molecules is immobilised onto a sensor chip and the other molecule is allowed to flow over the functionalised sensor chip. If binding occurs between the analyte and immobilised ligand, a measurable response will be generated. Whereas the BCA assay can be used to determine the total protein content, SPR and ELISA are used to determine the VEGF binding and active protein concentration of bevacizumab. Bakri et al³³ studied the VEGF binding of bevacizumab stored in a vial and syringe over six months time using ELISA. In our study. Biacore was used to study active protein concentration and VEGF binding of bevacizumab stored in vial and PC syringe for a six month time period. A decrease in antibody binding will cause a decrease in relative response unit (RU) that is generated. Biacore is a real time based method and is more sensitive compared to ELISA while no labelling is required.

Our results using several analytical methods and real time VEGF binding technique (Biacore) demonstrate that the commercial solution of bevacizumab (25 mg/mL, 16 mL in vial) can be fractionated in polypropylene and polycarbonate syringes and stored up to 6 months at 4°C without any changes in protein physical stability and VEGF binding of bevacizumab.

326 **Summary** 327 What was known before 328 Previous studies have revealed that repackaged bevacizumab in single use syringe and 329 ranibizumab have comparable outcome in terms of improvement of visual acuity for 330 AMD. Different studies on repackaged bevacizumab have been performed to evaluate 331 physicochemical stability in polypropylene syringe, with shelf storage of up to 3 332 months. The impact of storage of bevacizumab fractionated into PC polycarbonate 333 syringe for a longer period of time (6 months) on the quality of VEGF binding and 334 protein stability has not been determined. 335 What this study adds 336 This 6-month study indicates that the quality of bevacizumab repackaged into 337 prefilled PC polycarbonate syringes is not different from bevacizumab from a freshly 338 opened vial. As far as can be determined by SPR, the VEGF binding of bevacizumab 339 in the polycarbonate PC syringe was the same as that for bevacizumab taken from a 340 freshly opened vial. 341 Acknowledgements. HK, GS, PTK and SB are grateful for funding from NIHR 342 Biomedical Research Centre at Moorfields Hospital and the UCL Institute of 343 Ophthalmology, Moorfields Special Trustees, the Helen Hamlyn Trust (in memory of 344 Paul Hamlyn), Fight for Sight, Freemasons Grand Charity and Michael and Ilsa Katz 345 charity. GS and SB are also grateful for funding from the UK Engineering & Physical 346 Sciences Research Council (EPSRC) for the EPSRC Centre for Innovative 347 Manufacturing in Emergent Macromolecular Therapies. Financial support from the 348 consortium of industrial and governmental users for the EPSRC Centre is also 349 acknowledged.

Conflict of Interest. AF is an employee of Moorfileds Pharmaceuticals. The other
 authors declare that they have no conflict of interest.

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355	Figure 1. A; SDS-PAGE analysis of bevacizumab solution from the syringes (PC and		
356	PP) and vial, at T0, T3 and T6. Novex Bis-Tris 4-12% gels were stained with		
357	colloidal blue. Lane M: Protein standards. Lanes 1, 4, 7; bevacizumab (1.25 mg/ml)		
358	from vial at T0, T3 and T6 respectively. Lanes 2, 5, 8; bevacizumab (1.25 mg/ml)		
359	from PC syringes at T0, T3 and T6 respectively. Lanes 3, 6, 9; bevacizumab (1.25		
360	mg/ml) from PP syringes at T0, T3 and T6 respectively. B; Size exclusion		
361	chromatograms of bevacizumab from the PC syringe. C; Average percentage AUC		
362	for SEC peak at 58-59 minutes for bevacizumab solution from the PP and PC syringes		
363	and vial at T0, T3, T6 (n=3), No significant difference ($p > 0.05$) between presence of		
364	HMWS in vial and syringe over 6 months of storage. D; Overlay of size distribution		
365	curves for PC syringe and vial after six-month storage, bevacizumab solution from		
366	vial and syringe have a similar size distribution. E; DLS measurements of		
367	bevacizumab from PC syringes and vial at T0, T3 and T6 at 25 °C ($p > 0.05$).		
368			
369	Figure 2. A; The representative binding chart for bevacizumab in PC syringe at T6		
370	(N=3) and freshly opened vial at 1.25 and 0.625 mg/mL concentration, B; Binding		
371	sensograms of PC syringe at 1.25 mg/mL at T6 overlaid with bevacizumab from		
372	freshly opened vial, C; Biacore calculation of the active protein concentrations;		
373	bevacizumab obtained from syringes and the vial (N=3) at T0 and T6.		
374			
375	Figure 3. SDS-PAGE analysis of bevacizumab fractions eluted from SEC. Novex		
376	Bis-Tris 4-12% gels were stained with silver stain. Lane M: Protein standards. Lane 1:		
377	Bevacizumab from vial. Lane 2-4: Bevacizumab fractions at 58-60 minutes from SEC		
378	represent dimer of bevacizumab. Lane 5,6: Bevacizumab fractions at 71-72 minutes		
379	from SEC represent the monomer content.		
380 381			

383 Tables

Table 1

Purpose of study	Duration of study	Syringe material	Ref
Compare quality of repackaged bevacizumab from 5 different compounding pharmacies in UK	14 days	Polypropylene syringe	13
Examine the effect of silicon oil microdroplets and mishandling on protein aggregation level in repackaged bevacizumab	3 months	Plastic syringe (material not specified)	14
High molecular weight aggregates in repackaged bevacizumab	Not specified	Plastic syringe (material not specified)	24
Stability of bevacizumab repackaged in 1 mL polypropylene syringes for intravitreal administration	3 months	Polypropylene syringe	30

Table 1. Example studies of storage stability of repackaged bevacizumab in syringes.

Table 2

Compositions	Polycarbonate (PC)	Polypropylene (PP)
Barrel	Luer-lock	Slip-lock
Plunger Rod	Polypropylene	Polypropylene
Stopper	Latex free elastomer	Polyisoprene
Lubricant	Silicone	Silicone
Sterilisation Method	Gamma irradiation	Gamma irradiation
Supplier	B. Braun Medical Inc	Becton Dickinson
	(Cat. No 309628)	(Cat. No 9161406V)

Table 2. Material compositions for Polycarbonate (PC) and Polypropylene (PP) syringes.

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