



The Classic

Review of Dohan Eherenfest et al. (2009) on “Classification of platelet concentrates: From pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF)”



Theodorakys Marín Fermín^{a,b,1,*}, Jacob G. Calcei^{c,1}, Franco Della Vedova^{d,1},
 Juan Pablo Martínez Cano^{e,1}, Claudia Arias Calderon^{f,1}, Mohamed A. Imam^{g,h,1},
 Miguel Khoury^b, Markus W. Laupheimer^b, Pieter D'Hooghe^{b,1}

^a Centro Médico Profesional Las Mercedes, Las Mercedes 1060, Caracas, Venezuela

^b Aspetar Orthopaedic and Sports Medicine Hospital, Sports City Street, Inside Aspire Zone, Al Buwairda St, 29222, Doha, Qatar

^c University Hospitals Drusinsky Sports Medicine Institute, Case Western Reserve University, 11100 Euclid Ave, Cleveland, OH, 44106, USA

^d Instituto “Dr. Jaime Slullitel”, 2000 Rosario, Santa Fe, Argentina

^e Departamento de Ortopedia, Fundación Valle Del Lili, Cra 98 No. 18-49, 760032, Cali, Colombia

^f Department of Trauma and Orthopedic Surgery, Hospital Nacional Edgardo Rebagliati Martins, Jesús María 15072, Peru

^g Smart Health Centre, University of East London, London E16 2RD, UK

^h Rowley Bristow Orthopaedic Unit, Ashford and St Peter's Hospital, Ashford TW15 3AA, UK

ARTICLE INFO

Keywords:

Classification
 Growth factors
 Leucocytes
 Orthobiologics
 Platelet-rich plasma
 Platelet concentrates

ABSTRACT

This classic discusses the original publication of Dohan Eherenfest et al. on “Classification of platelet concentrates: from pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF)”, in which the authors propose four categories of platelet concentrates depending on their leucocyte and fibrin content (P-PRP, leucocyte- and platelet-rich plasma (L-PRP), pure platelet-rich fibrin (P-PRF), and L-PRF) to group a “jungle” of products in which the term platelet-rich plasma (PRP) was used indistinctly. They were able to identify common factors such as: (1) the use of anticoagulants and immediate centrifugation of the blood after its collection; (2) most preparation techniques allowed platelet concentrate preparation within an hour; (3) the centrifugation aimed to separate the blood in layers that would allow the extraction of specific fractions; and (4) the product was activated with thrombin or calcium chloride. The reviewed manuscript has been listed among the most cited PRP articles in regenerative medicine, with more than 800 citations, driving current scientific research and clinical practise by categorising L-PRP and P-PRP (now, leucocyte-poor PRP). The classification has also opened the door to understanding intrinsic biological mechanisms between platelets, leukocytes, fibrin, and growth factors, which will later be considered for studying the proliferation and differentiation of cells in different tissues affected by PRP. Since the initial classification of platelet concentrates, several other classification systems have been proposed and published in the current literature such as platelet, activation, white blood cell (PAW), Mishra, platelet, leucocyte, red blood cells, and activation (PLRA), dose of platelet, efficiency, purity, and activation (DEPA), method, activation, red blood cells, spin, platelets, image guidance, leukocytes, and light activation (MARSPILL), etc. These classifications have identified important aspects of PRP that affect the biological composition and, ultimately, the indications and outcomes. To date, there is still a lack of standardisation in sample preparation, cohort heterogeneity, and incomplete reporting of sample preparation utilised, leading to a lack of clarity and challenging researchers and clinicians.

Abbreviations: DEPA, Dose of platelet, efficiency, purity, and activation; L-PRF, Leucocyte- and platelet-rich fibrin; L-PRP, Leucocyte- and platelet-rich plasma; MARSPILL, Method, activation, red blood cells, spin, platelets, image guidance, leukocytes, and light activation; MIBO, Minimum information for studies evaluating biologics in orthopaedics; PAW, Platelet, activation, white blood cell; PLRA, Platelet, leucocyte, red blood cells, and activation; P-PRF, Pure platelet-rich fibrin; P-PRP, Pure platelet-rich plasma (now, leucocyte-poor platelet-rich plasma); PRP, Platelet-rich plasma.

* Corresponding author. Centro Médico Profesional Las Mercedes, Las Mercedes, Caracas 1060, Venezuela. Tel.: +58 4143727046.

E-mail address: theodorakysmarin@yahoo.com (T. Marín Fermín).

¹ ISAKOS Young Professionals Task Force.

<https://doi.org/10.1016/j.jisako.2023.07.010>

Received 6 December 2022; Received in revised form 28 June 2023; Accepted 27 July 2023

Available online 8 August 2023

2059-7754/© 2023 The Author(s). Published by Elsevier Inc. on behalf of International Society of Arthroscopy, Knee Surgery and Orthopedic Sports Medicine. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

INTRODUCTION

Platelet-rich plasma (PRP) is a worldwide-implemented regenerative medicine therapy. The clinical applications and use of platelet-rich therapy in medicine and surgery have thrived over the past two decades. It was described in 1970 as a plasma portion from autologous blood with increased platelet concentration obtained by a centrifugation process [1].

PRP harnesses the signalling molecules and growth factors of platelets such as vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), epidermal growth factor, insulin-like growth factors (e.g., IGF-1, IGF-2), and others that enhance the natural healing potential of tissues, pain and inflammation modulation, and functional improvement [1].

PRP is currently used to treat multiple musculoskeletal conditions, including knee osteoarthritis, cartilage injuries, patellar tendonitis, and tennis elbow [2,3]. The media, celebrity athletes, the desire for novel treatments, and its autologous nature have boosted its use in multiple sports-related injuries during the last decades [4–7]. As a result, its high demand has led to the industry-driven development of various platelet concentrate systems and products, exceeding the pace of evidence-based practise [1,3,7,8].

In 2009, Dohan Ehrenfest et al. proposed the first attempt at PRP classification to categorise the platelet concentrates concerning their fibrin and leucocyte content and the degree of standardization of the procedure, providing an overview of the available systems [9]. This classification later inspired authors to investigate the role of the different PRP components and the development of new classifications and reporting guidelines, highlighting leucocyte properties in orthobiologic products. However, the goal of standardisation still seems far away. A systematic review by Magalon et al. [8] revealed great heterogeneity among fifty platelet concentrate products from forty companies, which may explain the inconsistent outcomes in the literature. Thus, as a scientific orthopaedic community, we should question ourselves: how far have we gotten from the first attempt at orthobiologic classification? or has the panorama changed since then?

CONSIDERATION

Historical perspective

In 2009, Dohan Ehrenfest et al. [9] faced a scenario where commercial interests were obscuring real clinical benefits, a “jungle” in their own words, developing a plethora of preparation methods, systems, and centrifuges, and multiple platelet-derived products were covered by the umbrella term PRP, which did not allow a distinction between them. However, the authors were able to identify common factors among the available products such as: (1) the use of anticoagulants and immediate centrifugation of the blood after its collection; (2) most preparation techniques allowed platelet concentrate preparation within an hour (3) the centrifugation aimed to separate the blood in layers that would allow the extraction of specific fractions; and (4) the product was activated with thrombin or calcium chloride. The situation led his team to propose a classification to provide an objective approach to the growth and advancement of PRP therapy [9].

Back then, Dohan Ehrenfest et al. were implementing Choukroun's leucocyte- and platelet-rich fibrin (L-PRF) protocol in oral and maxillo-facial surgery (Box 1 and 2) [10]. The technique's benefits included a high efficiency in retrieving and concentrating platelets and leukocytes and a semisolid and three-dimensional fibrin matrix structure mimicking a natural blood clot. However, it was technically demanding because its success depended on rapid blood collection and centrifugation. After all, the lack of anticoagulant deemed almost instantaneous coagulation of the blood once in contact with the walls of the dry-glass tubes. Otherwise, the fibrin would polymerise diffusely in the tube, failing to concentrate most of the available platelets.

Understanding the rise of the idea: the clinical implication

The proposed platelet concentrate classification of Dohan Ehrenfest et al. included three main parameters (Table 1), allowing the characterisation of platelet concentrates in pure platelet-rich plasma (P-PRP), leucocyte- and platelet-rich plasma (L-PRP), pure platelet-rich fibrin (P-PRF), and L-PRF [9].

This classification brought new insights into the relevance of other components of platelet concentrates, the leukocytes, and fibrin. By then, anti-microbial and immunomodulatory properties were already attributed to the leukocytes in platelet concentrates and their role in angiogenesis with vascular endothelial growth factor synthesis [11–14]. The role of L-PRP in tendon healing was starting to rise [15]. Similarly, they supported the theory that the fibrin matrix and its composition (including cytokines) were crucial for the platelet concentrate's clinical efficacy. In fact, the clotting pattern they were inducing in Choukroun's L-PRF technique enhanced platelet growth factor release. The author confirmed his claims with the publication of another article in 2012 [16], where his team proved not only an increased release of growth factors but also a more extended release period (seven days) due to the naturally formed dense fibrin network in contrast to the light fibrin network present in P-PRP, observed when artificial activation was triggered with bovine thrombin, calcium chloride, or other clotting agents [17–19].

Scientific and societal impact

In a recent publication by the European Society for Sports Traumatology, Knee Surgery, and Arthroscopy Orthobiologic Initiative, the paper by Dohan Ehrenfest et al. [9] was listed fourth among the most cited PRP articles in regenerative medicine with more than 800 citations, including two additional papers from the author in the top 100 [6]. The current scientific research and clinical practise, driven by the main categorisation between L-PRP and P-PRP (now, leucocyte-poor PRP), confirm the impact of this article. Indeed, recent studies have proposed specific roles for different leukocytes in PRP clinical efficiency depending on the healing stage and the type of injury [20]. Lymphocytes, for example, have an anti-inflammatory role by steering monocyte differentiation from the M1 to the M2 subtype. On the other hand, neutrophils lead the so-called “regenerative inflammation” by secreting chemokines to recruit macrophages and promoting an inflammatory process desired to trigger the healing process [20,21].

BOX 1

Choukroun's L-PRF protocol [10].

This novel method was characterised by its simplicity, reproducibility, and low cost. In this protocol, developed in Nice (France), venous blood was collected in 10 ml tubes and instantly centrifuged without anticoagulant. The lack of an anticoagulant allowed the natural formation of a clot (platelet activation) that would facilitate the manipulation of L-PRF, avoiding the implementation of any additive. Thus, after centrifugation at 3000 rpm (400g) for 10 min, three distinct layers are visualised: the red blood cell layer at the bottom of the tube, a top acellular plasma layer, and a L-PRF clot in between, containing most of the platelets.

BOX 2

Short interview with Pr. Lars Rasmusson – Co-author of the classic paper on “Classification of platelet concentrates: from pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF)”

Head of Department, Department of Maxillofacial Surgery, Sahlgrenska Academy, University of Gothenburg.

Q1: What motivated the development of the classification system?

A-LR: The motivation to develop a classification system was the somewhat conflicting PRP and PRF handling methodologies at the time.

Q2: Who were the researchers involved in the development of the classification and their contribution to it?

A-LR: It was Pr. Tomas Albrektsson, Head of the Department of Biomaterials, University of Gothenburg, and I at the time, together with our postdoc, David Dohan Ehrenfest.

Q3: What were the clinical uses of PRP in your Institution?

A-LR: PRP and PRF were used both clinically in maxillofacial reconstruction and in experimental work at our lab. In clinical practise here at our unit, L-PRF has replaced PRP since it is easier to use and possible to manufacture in different consistency/preparations, for example, injectable and as membranes.

Q4: What are the current challenges of PRP therapy?

A-LR: The challenge has been (and still is) to prove long-term efficacy and superiority in bone healing.

Q5: What is the future of PRP therapy?

A-LR: I strongly believe that more indications for use will be discovered and evaluated in cartilage repair via injectable platelets.

Before the classification system proposed by Dohan Ehrenfest et al. [9], the characterization of platelet concentrates was confusing and contradictory and created a methodological bias in many publications. The authors addressed this challenging problem by simply proposing a classification framework. Additionally, the classification opened the door to understanding intrinsic biological mechanisms between the platelets, leukocytes, fibrin, and growth factors later considered for studying the proliferation and differentiation of cells in different tissues affected by PRP. In other words, the authors proposed evaluating these products as living tissues instead of pharmaceutical preparations with a simple and precise composition.

Current evidence

After highlighting the importance of classifying the platelet concentrates according to the presence of leukocytes, several clinical and laboratory studies conducted during the last decade have demonstrated clinical benefit, especially in tendinopathies. In a controlled laboratory study, Lin et al. [22] revealed a higher induction of platelet growth

factors and tenocyte proliferation with L-PRP preparations than P-PRP. Furthermore, a network meta-analysis of 18 randomised controlled trials by Fitzpatrick et al. [3] showed that the most significant positive outcomes were obtained from a single ultrasound-guided L-PRP injection in tendinopathies such as the rotator cuff, tennis elbow, patellar tendon, and Achilles tendon.

On the other hand, the presence of leukocytes in PRP has been found to be chondrotoxic, while in the absence of leukocytes, PRP promotes chondrogenesis [23,24]. However, clinical studies on knee osteoarthritis have shown conflicting results when comparing both PRP preparations [24–26]. Although L-PRP has offered comparable results to leucocyte-poor preparations, the latter is preferred due to a higher risk of swelling from the increased inflammatory response [27].

Lessons learnt

As with many biologic therapies in medicine, particularly musculoskeletal medicine, additional research has often led to more questions than answers. Quite possibly the most important lesson learnt with

BOX 3

Summary of The Classic

PRP is currently used to treat multiple musculoskeletal conditions, including knee osteoarthritis, cartilage injuries, patellar tendonitis, and tennis elbow. In recent decades, the media, celebrity athletes, the desire for novel treatments, and its autologous nature have boosted its use in multiple sports-related injuries. As a result, its high demand has led to the industry-driven development of various platelet concentrate systems and products, exceeding the pace of evidence-based practise.

In 2009, Dohan Ehrenfest et al. proposed the first attempt at PRP classification to categorise the platelet concentrates concerning their fibrin and leucocyte content and the degree of standardisation of the procedure, providing an overview of the available systems. The authors proposed four categories of platelet concentrates (P-PRP, L-PRP, P-PRF, and L-PRF) to group a “jungle” of products in which the term PRP was used indistinctly.

The classification opened the door to understanding intrinsic biological mechanisms between the PRP components. Since the initial classification, several other classification systems have been proposed and published in the current literature, identifying important aspects of PRP. To date, there is still a lack of standardisation in sample preparation, cohort heterogeneity, and incomplete reporting of sample preparation utilised, leading to a lack of clarity and challenging researchers and clinicians. A paucity of accurate reporting of a highly variable product has led to a lack of clarity and continues to challenge researchers and clinicians. In this sense, journal editors can play an essential role in evidence quality improvement by requesting mandatory adherence to acceptable orthobiologic reporting guidelines in submission and review. The goal of the present decade is to build a new body of evidence with high-quality reporting and reproducibility that will serve as the foundation of its long-awaited standardisation.

Table 1
Dohan Ehrenfest et al. (2009) platelet concentrate classification.

PARAMETERS	Subparameter
A Preparation kits and centrifuges	1 Size of the centrifuge
	2 Duration of the procedure
	3 Cost of the device and kits
	4 Ergonomics of the kit and the complexity of the procedure
B Content of the concentrate	1 Final volume of usable concentrate
	2 Efficiency in collecting platelets
	3 Leucocytes
	4 Preservation of the components
C Fibrin network	1 Concentration and density
	2 Polymerization process

biologic treatments, and PRP specifically, is that there is a vast range of variability not only in the PRP preparation (instruments, devices, spin rate and time, activators, among others) but also in the quality of the product due to inter- and intra-human variability. Since PRP is an autologous product, the quality of the sample, growth factor concentration, and activity of the components within the specimen are likely affected by the health status of the individual, medications, diet, and cortisol stress levels, among others. Furthermore, the heterogeneity of clinical outcomes among the studies has also moved the spotlight to the key elements that allow patients to benefit from PRP therapy. Researchers have now classified patients into responders and non-responders and started phenotyping the ideal patient [28–31].

New developments: new classifications and research originated from the original study

Since the initial classification of platelet concentrates, several other classification systems have been proposed and published in the current literature. Rossi et al. [32] recently reviewed the available classification systems for PRP, examining the advantages and limitations of each (Fig. 1). Rossi et al. acknowledge the Dohan Ehrenfest classification system and its basic component breakdown.

DeLong et al. [33] presented their classification system in 2012, known as the platelet, activation, white blood cell (PAW) classification. The PAW classification system was based on the absolute number of platelets (P1 to P4, depending on the number of platelets), the method of platelet activation, and the presence (α) or absence (β) of white blood cells. Mishra et al. [34] presented a similar classification system. However, they classified the variables differently, which resulted in four

separate categories of PRP: elevated platelets and leukocytes without an external activator, elevated platelets and leukocytes with an external activator, elevated platelets without leukocytes and no external activator, and elevated platelets without leukocytes but with an external activator.

The classification systems continued to evolve as Mautner et al. [35] noted the importance of red blood cells and their potential detrimental effects in PRP; thus, they added red blood cell analysis to the classification known as platelet, leucocyte, red blood cells, and activation (PLRA). Magalon et al. [36] then proposed the dose of platelet, efficiency, purity, and activation (DEPA) classification. In this classification system, the proportion of the platelets recovered from PRP and the purity of the PRP sample were included as essential qualities of the sample preparation. Lana et al. [37] proposed a classification in 2017 evaluating method, activation, red blood cells, spin, platelets, image guidance, leukocytes, and light activation (MARSPILL). Finally, The Platelet Physiology Subcommittee of the Scientific and Standardisation Committee of the International Society on Thrombosis and Haemostasis has recently proposed a classification system that includes the presence of leukocytes, red blood cells, activation products, platelet concentration, and preparation categories [38]. Individually, these classification systems have identified important aspects of PRP that affect the biological composition and, ultimately, the indications and outcomes. In any situation, simple and elegant classification systems are often preferred due to their ease of use; however, bulkier classification systems are typically more comprehensive. Based on the current indications for PRP, the presence or absence of leukocytes seems to be the main dividing point. Still, the complexity of the makeup of PRP makes a simple and elegant classification system quite challenging to develop. While there is no perfect classification system to rely entirely on when evaluating biologics, including PRP, the field strives to characterise this biological product better. Until we determine exactly what elements of PRP are most important in affecting outcomes, we may be stuck relying on bulky systems or even combining multiple classification systems.

Future directions

Much of the challenge in practising evidence-based medicine in the case of biological treatments is the lack of standardisation in sample preparation, the heterogeneity in cohorts, and the incomplete reporting of sample preparation utilised. Thus, there has been a call for minimum reporting standards for studies involving biologics in musculoskeletal care [39–42]. Specifically, Murray et al. [41] reported the 23-item checklist compiled by the PRP working group to report minimum

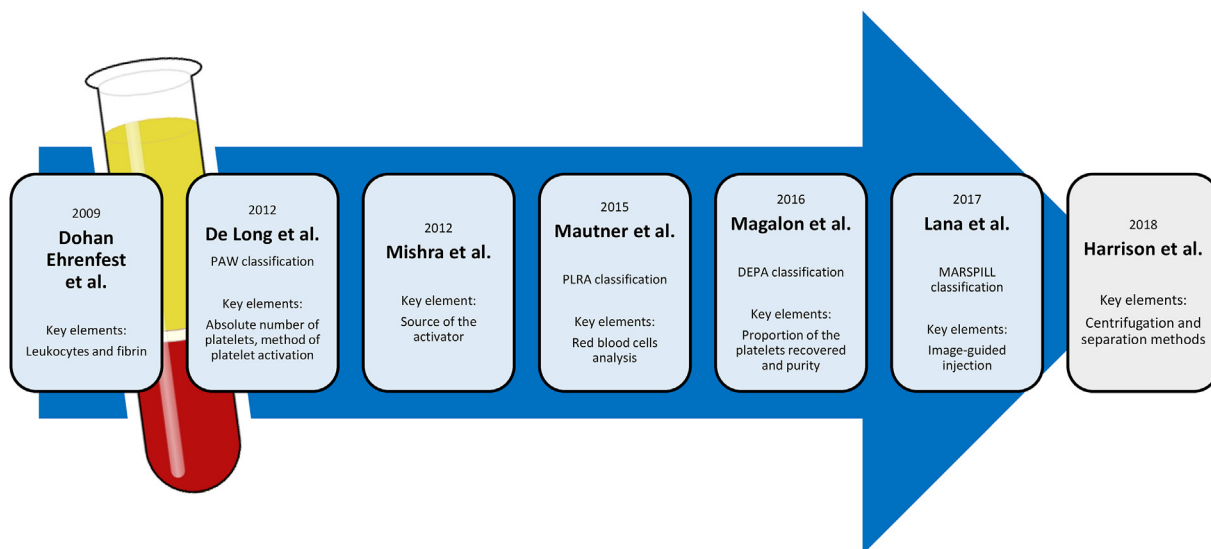


Fig. 1. Historical landmarks of platelet-rich plasma classifications.

information for studies evaluating biologics in orthopaedics (MIBO). In an assessment of the 50 most cited articles related to PRP in musculoskeletal medicine, Bugarin et al. [43] reported a high level of evidence in approximately 50% of the studies. Still, most of the studies were of only fair methodological quality. Systematic reviews by Chahla et al. [44], DeClercq et al. [45], and Marín Fermín et al. [5] have revealed that less than 10% of the studies provided a clear description of the implemented PRP preparation protocol, which significantly limits the study's reproducibility. A paucity of accurate reporting of a highly variable product has led to a lack of clarity and continues to challenge researchers and clinicians. In this sense, journal editors can play an essential role in evidence quality improvement by requesting mandatory adherence to acceptable orthobiologic reporting guidelines in submission and review [46]. The goal of the present decade is to build a new body of evidence with high-quality reporting and reproducibility that will serve as the foundation of its long-awaited standardisation.

ADDITIONAL EXPERT OPINION

Importantly, when approaching a novel treatment option, we must do our best to practise evidence-based medicine. This first requires research-driven processes to identify the crucial components of the product so that it can be appropriately characterised and reliably recreated. In the case of biologics such as PRP, where countless variables may ultimately impact the preparation and final product, it becomes vital to identify the key elements and differences in the preparation method. Classification systems can play a key role in driving this standardisation process. As mentioned above, the available classification systems have identified essential aspects of PRP that affect the biological composition and, ultimately, the indications and outcomes. The simple and elegant system proposed by Dohan Ehrenfest et al. [9] identified what seems to be the main dividing point based on the current indications for PRP in musculoskeletal medicine: the presence or absence of leukocytes. While there is no perfect classification system to rely entirely on when evaluating PRP, there is no question that the Dohan Ehrenfest classification system began the conversation, leading to many more comprehensive classification systems.

Contributions

TMF: conceptualization, investigation, writing original draft, visualization, project administration; JGC: investigation, writing - original draft; FDV: resources, writing - review & editing; JPMC: resources, writing - review & editing; CAC: resources, writing - review & editing; MI: writing - review & editing, supervision; MK: validation, data curation, writing - review & editing, supervision; MWL: validation, writing - review & editing, supervision; PDH: validation, writing - review & editing, supervision.

Funding

None.

Data availability statement

The data underlying this article are available in the article and in its online supplementary material.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgments

We thank Pr. Lars Rasmusson for his kind collaboration with the short interview included in this manuscript.

References

- [1] Obana KK, Schallmo MS, Hong IS, Ahmad CS, Moorman 3rd CT, Trofa DP, et al. Current trends in orthobiologics: an 11-year review of the orthopaedic literature. *Am J Sports Med* 2022 Sep;50(11):3121–9. <https://doi.org/10.1177/03635465211037343>.
- [2] Bansal H, Leon J, Pont JL, Wilson DA, Bansal A, Agarwal D, et al. Platelet-rich plasma (PRP) in osteoarthritis (OA) knee: correct dose critical for long term clinical efficacy. *Sci Rep* 2021 Feb 17;11(1):3971. <https://doi.org/10.1038/s41598-021-83025-2>.
- [3] Fitzpatrick J, Bulsara M, Zheng MH. The effectiveness of platelet-rich plasma in the treatment of tendinopathy: a meta-analysis of randomized controlled clinical trials. *Am J Sports Med* 2017 Jan;45(1):226–33. <https://doi.org/10.1177/0363546516643716>.
- [4] Makaram NS, Murray IR, Rodeo SA, Sherman SL, Murray AD, Haddad FS, et al. The use of biologics in professional and Olympic sport: a scoping review protocol. *Bone Jt Open* 2020 Nov 17;1(11):715–9. <https://doi.org/10.1302/2633-1462.111.BJO-2020-0159>.
- [5] Marín Fermín T, Papakostas E, Macchiarola L, Zampeli F, Kalifis G, De Girolamo L, et al. Injectable orthobiologics in professional football (Soccer) players: a systematic review. *J Cartilage Joint Preservation* 2022;2(2):100050. <https://doi.org/10.1016/j.jcjp.2022.100050>.
- [6] Coulange Zavarro A, De Girolamo L, Laver L, Sánchez M, Tischer T, Filardo G, et al. The top 100 most cited articles on platelet-rich plasma use in regenerative medicine-A bibliometric analysis-from the ESSKA orthobiologic initiative. *Bioengineering (Basel)* 2022 Oct 19;9(10):580. <https://doi.org/10.3390/bioengineering9100580>.
- [7] Frank RM, Sherman SL, Chahla J, Drago J, Mandelbaum B, Members of the Biologic Association, Anz AW, et al. Biologic association annual summit: 2020 report. *Orthop J Sports Med* 2021 Jun 7;9(6):23259671211015667. <https://doi.org/10.1177/23259671211015667>.
- [8] Magalon J, Brandin T, Francois P, Degioanni C, De Maria L, Grimaud F, et al. Technical and biological review of authorized medical devices for platelets-rich plasma preparation in the field of regenerative medicine. *Platelets* 2021 Feb 17;32(2):200–8. <https://doi.org/10.1080/09537104.2020.1832653>.
- [9] Dohan Ehrenfest DM, Rasmusson L, Albrektsson T. Classification of platelet concentrates: from pure platelet-rich plasma (P-PRP) to leukocyte- and platelet-rich fibrin (L-PRF). *Trends Biotechnol* 2009 Mar;27(3):158–67. <https://doi.org/10.1016/j.tibtech.2008.11.009>.
- [10] Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhyi J, et al. Platelet-rich fibrin (PRF): a second-generation platelet concentrate. Part I: technological concepts and evolution. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006 Mar;101(3):e37–44. <https://doi.org/10.1016/j.tripleo.2005.07.008>.
- [11] Cieslik-Bielecka A, Gazdzik TS, Bielecki TM, Cieslik T. Why the platelet-rich gel has anti-microbial activity? *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2007 Mar;103(3):303–5. <https://doi.org/10.1016/j.tripleo.2006.08.034>. author reply 305–6.
- [12] Moojen DJ, Everts PA, Schure RM, Overvest EP, van Zundert A, Knappe JT, et al. Anti-microbial activity of platelet-leukocyte gel against *Staphylococcus aureus*. *J Orthop Res* 2008 Mar;26(3):404–10.
- [13] El-Sharkawy H, Kantarci A, Deady J, Hasturk H, Liu H, Alshahat M, et al. Platelet-rich plasma: growth factors and pro- and anti-inflammatory properties. *J Periodontol* 2007 Apr;78(4):661–9. <https://doi.org/10.1902/jop.2007.060302>.
- [14] Werther K, Christensen IJ, Nielsen HJ. Determination of vascular endothelial growth factor (VEGF) in circulating blood: significance of VEGF in various leucocytes and platelets. *Scand J Clin Lab Invest* 2002;62(5):343–50. <https://doi.org/10.1080/00365510260296492>.
- [15] Schnabel LV, Mohammed HO, Miller BJ, McDermott WG, Jacobson MS, Santangelo KS, et al. Platelet rich plasma (PRP) enhances anabolic gene expression patterns in flexor digitorum superficialis tendons. *J Orthop Res* 2007 Feb;25(2):230–40. <https://doi.org/10.1002/jor.20278>.
- [16] Dohan Ehrenfest DM, Bielecki T, Jimbo R, Barbé G, Del Corso M, Inchingolo F, et al. Do the fibrin architecture and leukocyte content influence the growth factor release of platelet concentrates? An evidence-based answer comparing a pure platelet-rich plasma (P-PRP) gel and a leukocyte- and platelet-rich fibrin (L-PRF). *Curr Pharmaceut Biotechnol* 2012 Jun;13(7):1145–52. <https://doi.org/10.2174/138920112800624382>.
- [17] Dohan Ehrenfest DM, Del Corso M, Inchingolo F, Sammartino G, Charrier JB. Platelet-rich plasma (PRP) and platelet-rich fibrin (PRF) in human cell cultures: growth factor release and contradictory results. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2010 Oct;110(4):418–21. <https://doi.org/10.1016/j.tripleo.2010.05.059>. author reply 421–2.
- [18] Fernández-Barbero JE, Galindo-Moreno P, Avila-Ortiz G, Caba O, Sánchez-Fernández E, Wang HL. Flow cytometric and morphological characterization of platelet-rich plasma gel. *Clin Oral Implants Res* 2006 Dec;17(6):687–93. <https://doi.org/10.1111/j.1600-0501.2006.01179.x>.
- [19] Mazzucco L, Balbo V, Cattana E, Guaschino R, Borzini P. Not every PRP-gel is born equal. Evaluation of growth factor availability for tissues through four PRP-gel preparations: fibrinet, RegenPRP-Kit, Plateltex and one manual procedure. *Vox Sang* 2009 Aug;97(2):110–8. <https://doi.org/10.1111/j.1423-0410.2009.01188.x>.
- [20] Lana JF, Huber SC, Purita J, Tambeli CH, Santos GS, Paulus C, et al. Leukocyte-rich PRP versus leukocyte-poor PRP - the role of monocyte/macrophage function in the healing cascade. *J Clin Orthop Trauma* 2019 Oct;10(Suppl 1):S7–12. <https://doi.org/10.1016/j.jcot.2019.05.008>.
- [21] Marathe A, Patel SJ, Song B, Slipek JM, Shybut TS, Lee BH, et al. Double-spin leukocyte-rich platelet-rich plasma is predominantly lymphocyte rich with notable

- concentrations of other white blood cell subtypes. *Arthrosc Sports Med Rehabil* 2021 Dec 10;4(2):e335–41. <https://doi.org/10.1016/j.asmr.2021.10.004>.
- [22] Lin KY, Chen P, Chen AC, Chan YS, Lei KF, Chiu CH. Leukocyte-rich platelet-rich plasma has better stimulating effects on tenocyte proliferation compared with leukocyte-poor platelet-rich plasma. *Orthop J Sports Med* 2022 Mar 15;10(3):23259671221084706. <https://doi.org/10.1177/23259671221084706>.
- [23] Xu Z, Yin W, Zhang Y, Qi X, Chen Y, Xie X, et al. Comparative evaluation of leukocyte- and platelet-rich plasma and pure platelet-rich plasma for cartilage regeneration. *Sci Rep* 2017 Mar 7;7:43301. <https://doi.org/10.1038/srep43301>.
- [24] Riboh JC, Saltzman BM, Yanke AB, Fortier L, Cole BJ. Effect of leukocyte concentration on the efficacy of platelet-rich plasma in the treatment of knee osteoarthritis. *Am J Sports Med* 2016 Mar;44(3):792–800. <https://doi.org/10.1177/0363546515580787>.
- [25] Di Martino A, Boffa A, Andriolo L, Romandini I, Altamura SA, Cenacchi A, et al. Leukocyte-rich versus leukocyte-poor platelet-rich plasma for the treatment of knee osteoarthritis: a double-blind randomized trial. *Am J Sports Med* 2022 Mar;50(3):609–17. <https://doi.org/10.1177/03635465211064303>.
- [26] Abbas A, Du JT, Dhotar HS. The effect of leukocyte concentration on platelet-rich plasma injections for knee osteoarthritis: a network meta-analysis. *J Bone Joint Surg Am* 2022 Mar 16;104(6):559–70. <https://doi.org/10.2106/JBJS.20.02258>.
- [27] Kim JH, Park YB, Ha CW, Roh YJ, Park JG. Adverse reactions and clinical outcomes for leukocyte-poor versus leukocyte-rich platelet-rich plasma in knee osteoarthritis: a systematic review and meta-analysis. *Orthop J Sports Med* 2021 Jun 30;9(6):23259671211011948. <https://doi.org/10.1177/23259671211011948>.
- [28] Mantripragada VP, Csorba A, Bova W, Boehm C, Piuuzzi NS, Bullen J, et al. Assessment of clinical, tissue, and cell-level metrics identify four biologically distinct knee osteoarthritis patient phenotypes. *Cartilage* 2022 Jan-Mar;13(1):19476035221074003. <https://doi.org/10.1177/19476035221074003>.
- [29] Ganguly P, Fiz N, Beitia M, Owston HE, Delgado D, Jones E, et al. Effect of combined intraosseous and intraarticular infiltrations of autologous platelet-rich plasma on subchondral bone marrow mesenchymal stromal cells from patients with hip osteoarthritis. *J Clin Med* 2022 Jul 4;11(13):3891. <https://doi.org/10.3390/jcm11133891>.
- [30] Zahir H, Dehghani B, Yuan X, Chinenov Y, Kim C, Burge A, et al. In vitro responses to platelet-rich-plasma are associated with variable clinical outcomes in patients with knee osteoarthritis. *Sci Rep* 2021 Jun 1;11(1):11493. <https://doi.org/10.1038/s41598-021-90174-x>.
- [31] Saita Y, Kobayashi Y, Nishio H, Wakayama T, Fukusato S, Uchino S, et al. Predictors of effectiveness of platelet-rich plasma therapy for knee osteoarthritis: a retrospective cohort study. *J Clin Med* 2021 Sep 29;10(19):4514. <https://doi.org/10.3390/jcm10194514>.
- [32] Rossi LA, Murray IR, Chu CR, Muschler GF, Rodeo SA, Piuuzzi NS. Classification systems for platelet-rich plasma. *Bone Joint Lett J* 2019 Aug;101-B(8):891–6. <https://doi.org/10.1302/0301-620X.101B8.BJJ-2019-0037.R1>.
- [33] DeLong JM, Russell RP, Mazzocca AD. Platelet-rich plasma: the PAW classification system. *Arthroscopy* 2012 Jul;28(7):998–1009. <https://doi.org/10.1016/j.arthro.2012.04.148>.
- [34] Mishra A, Harmon K, Woodall J, Vieira A. Sports medicine applications of platelet rich plasma. *Curr Pharmaceut Biotechnol* 2012 Jun;13(7):1185–95. <https://doi.org/10.2174/138920112800624283>.
- [35] Mautner K, Malanga GA, Smith J, Shiple B, Ibrahim V, Sampson S, et al. A call for a standard classification system for future biologic research: the rationale for new PRP nomenclature. *PM R* 2015 Apr;7(4 Suppl):S53–9. <https://doi.org/10.1016/j.pmrj.2015.02.005>.
- [36] Magalon J, Chateau AL, Bertrand B, Louis ML, Silvestre A, Giraudo L, et al. DEPA classification: a proposal for standardising PRP use and a retrospective application of available devices. *BMJ Open Sport Exerc Med* 2016 Feb 4;2(1):e000060. <https://doi.org/10.1136/bmjsem-2015-000060>.
- [37] Lana JFSD, Purita J, Paulus C, Huber SC, Rodrigues BL, Rodrigues AA, et al. Contributions for classification of platelet rich plasma - proposal of a new classification: MARSPILL. *Regen Med* 2017 Jul;12(5):565–74. <https://doi.org/10.2217/rme-2017-0042>.
- [38] Harrison P, Subcommittee on Platelet Physiology. The use of platelets in regenerative medicine and proposal for a new classification system: guidance from the SSC of the ISTH. *J Thromb Haemostasis* 2018 Sep;16(9):1895–900. <https://doi.org/10.1111/jth.14223>.
- [39] Rodeo S. The need for minimum reporting standards for studies of "biologics" in sports medicine. *Am J Sports Med* 2019 Sep;47(11):2531–2. <https://doi.org/10.1177/0363546519872219>.
- [40] LaPrade RF, Graden NR, Kahat DH. Editorial commentary: platelet-rich plasma: the devil is in the details, and the details need to be better reported. *Arthroscopy* 2019 Nov;35(11):3114–6. <https://doi.org/10.1016/j.arthro.2019.08.016>.
- [41] Murray IR, Geeslin AG, Goudie EB, Petrigliano FA, LaPrade RF. Minimum information for studies evaluating biologics in orthopaedics (MIBO): platelet-rich plasma and mesenchymal stem cells. *J Bone Joint Surg Am* 2017 May 17;99(10):809–19. <https://doi.org/10.2106/JBJS.16.00793>.
- [42] Murray IR, Murray AD, Geeslin AG, Goudie EB, White TO, Petrigliano FA, et al. Infographic: we need minimum reporting standards for biologics. *Br J Sports Med* 2019 Aug;53(15):974–5. <https://doi.org/10.1136/bjsports-2017-098122>.
- [43] Bugarin A, Schroeder G, Shi BY, Jones KJ, Kremen Jr TJ. Assessment of characteristics and methodological quality of the top 50 most cited articles on platelet-rich plasma in musculoskeletal medicine. *Orthop J Sports Med* 2022 May 26;10(5):23259671221093074. <https://doi.org/10.1177/23259671221093074>.
- [44] Chahla J, Cinque ME, Piuuzzi NS, Mannava S, Geeslin AG, Murray IR, et al. A call for standardization in platelet-rich plasma preparation protocols and composition reporting: a systematic review of the clinical orthopaedic literature. *J Bone Joint Surg Am* 2017 Oct 18;99(20):1769–79. <https://doi.org/10.2106/JBJS.16.01374>.
- [45] DeClercq MG, Fiorentino AM, Lengel HA, Ruzbarsky JJ, Robinson SK, Oberlohr VT, et al. Systematic review of platelet-rich plasma for rotator cuff repair: are we adhering to the minimum information for studies evaluating biologics in orthopaedics? *Orthop J Sports Med* 2021 Dec 7;9(12):23259671211041971. <https://doi.org/10.1177/23259671211041971>.
- [46] Marín Fermín T, Scarlat MM, Laupheimer MW. Would you have an injection without knowing its formula? New challenges in platelet-rich plasma therapy. *Int Orthop* 2022 Oct;46(10):2179–80. <https://doi.org/10.1007/s00264-022-05566-z>.